Informational Paper

The Pathogenesis of Periodontal Diseases

This informational paper was prepared by the Research, Science, and Therapy Committee of The American Academy of Periodontology, and is intended for the information of the dental profession. The purpose of the paper is to provide an overview of current knowledge relating to the pathogenesis of periodontal diseases. The paper will review biological processes thought to provide protection against periodontal infections. It will further discuss the mechanisms thought to be responsible for both overcoming and subverting such protective mechanisms and those that lead to destruction of periodontal tissues. Since an understanding of pathogenic mechanisms of disease is one foundation upon which new diagnostic and therapeutic modalities are based, the practitioner can use this information to help make decisions regarding the appropriate application of such new modalities in patient care settings. *J Periodontol* 1999;70:457-470.

athogenesis deals with the mode of origin or development of disease. In this paper, currently accepted concepts of the origin and progression of gingivitis and periodontitis are discussed. Since nearly all of the periodontal diseases are associated with and thought to be caused by microorganisms, some references to etiologic agents are of necessity utilized, particularly when certain disease processes are clarified by example.

Periodontal diseases comprise a variety of conditions affecting the health of the periodontium. Although the classification scheme defined at the 1989 World Workshop in Clinical Periodontics subdivided these diseases into a number of clinically defined subforms, 1 subsequent attempts to categorize patients according to the defined criteria have demonstrated the considerable problem of overlap in the disease definitions.² Furthermore, many of the microbiological and host response features of these diseases are common to several of the subforms of periodontitis. It has been the consensus of several groups, including the 1996 World Workshop in Periodontics,³ that the current classification scheme requires revision. Such a revision could lead to considerably improved diagnostic categories if the disease definitions were dependent upon knowledge of the etiology and pathogenesis of the various disease subforms as well as

upon more traditional parameters such as signs of inflammation, probing depths, clinical attachment loss, and age of onset.

Thus, although considerable progress has been made in defining both etiologic agents and pathways of pathogenesis in various forms of periodontal diseases, insufficient information exists to definitively recategorize these diseases. The approach to describing pathogenic mechanisms in this paper will, therefore, be in part generic and thus refer to "gingivitis" and "periodontitis" rather than to specific disease subforms. Where appropriate, descriptions of evidence for specific or unique pathways associated with specific forms of disease (as defined at the 1989 World Workshop in Clinical Periodontics) will be presented.

PATHOGENESIS OF GINGIVITIS

Chronic marginal gingivitis is characterized clinically by gingival redness, edema, bleeding, changes in contour, loss of tissue adaptation to the teeth, and increased flow of gingival crevicular fluid (GCF).^{4,5} Development of gingivitis requires the presence of plaque bacteria^{6,7} which are thought to induce pathological changes in the tissues by both direct and indirect means.⁸ Histopathologic observations have led to the subdivision of gingivitis into 3 stages.⁸⁻¹⁰ The initial lesion appears as an acute inflammatory response with characteristic infiltration with neutrophils. Vascular changes, epithelial cell changes, and collagen degradation are apparent. These initial changes are likely due to chemotac-

J Periodontol • April 1999 457

^{*} This paper was developed under the direction of the Committee on Research, Science and Therapy and approved by the Board of Trustees of The American Academy of Periodontology in January 1999.

tic attraction of neutrophils by bacterial constituents and direct vasodilatory effects of bacterial products, as well as activation of host systems such as the complement and kinin systems and arachidonic acid pathways. 11,12

The early lesion is characterized by a lymphoid cell infiltrate dominated by T lymphocytes, with extension of collagen loss, while the established lesion is dominated by B lymphocytes and plasma cells. Although direct evidence for specific mechanisms explaining the appearance and progression of gingivitis lesions is not available, the chronic inflammatory infiltrate characteristic of the early and established lesions, as well as the proliferation of the junctional epithelium and destruction of collagen, are consistent with the activation of mononuclear phagocytes and fibroblasts by bacterial products with the recruitment and activation of the local immune system and cytokine pathways. The progression of the lesion from acute inflammation through T cell and then B cell predominance is likely orchestrated by a progression of cytokines (dealt with in more detail below) which are responsible for recruitment, differentiation, and growth of the characteristic cell types with progressive chronicity of the lesion. Importantly, meticulous removal of plaque will usually result in resolution of the chronic gingivitis lesion without residual tissue destruction.

Acute necrotizing ulcerative gingivitis (ANUG), an acute infection of the gingiva characterized by interdental soft tissue necrosis and ulceration, pain, and bleeding, 13 is characterized histologically by frank invasion of the gingival connective tissues by spirochetes and a predominance of Prevotella intermedia and Fusobacterium nucleatum in the non-spirochetal flora. 13 The association of ANUG with recent episodes of stress, or with other conditions of impaired host defense such as malnutrition, immunosuppression, and systemic diseases, implicates any of a number of possible environmental and systemic stressors as pathogenic factors leading to the expression of the same syndrome. 14-20 A common feature of nearly all cases is very poor oral hygiene, and nearly all cases can be managed with local debridement, improved plague control, and judicious use of antibiotics.

Pathologic changes in the gingival tissues consistent with clinically chronic or acute gingivitis have been noted in a number of systemic condi-

tions.²¹⁻²³ Some of these conditions may mimic the vascular alterations seen in plaque-induced gingivitis or result in cellular infiltration by aberrant leukocytes or other vascular elements. These include acute leukemia, hemophilia, Sturge-Weber syndrome, and Wegener's granulomatosis. In other cases a defective host response to bacterial infection may be manifested as an overexpression of gingival inflammation or caused by an alteration in the usual bacterial microflora. Such conditions include Addison's disease, diabetes mellitus, thrombocytopenia, combined immunodeficiency diseases, and HIV infection. A third group of these conditions is related to hormonal changes manifested as an exaggerated inflammatory response to plaque as well as an alteration in the subgingival microflora. These include changes associated with pregnancy, puberty, steroid therapy, and use of birth control medications.²⁴⁻²⁷ Finally, a large number of drugs, many of which are associated with therapy for seizure disorders, hypertension, or transplant rejection, cause gingival enlargement in the presence of bacterial plaque. 28-33

PATHOGENESIS OF PERIODONTITIS

Periodontitis is clinically differentiated from gingivitis by the loss of the connective tissue attachment to the teeth in the presence of concurrent gingival inflammation.³⁴ Loss of the periodontal ligament and disruption of its attachment to cementum, as well as resorption of alveolar bone occurs. Together with loss of attachment, there is migration of the epithelial attachment along the root surface and resorption of bone.⁹ The histopathology of the periodontitis lesion is in many ways similar to that of the established lesion of gingivitis, with a predominance of plasma cells, loss of soft connective tissue elements, and, in addition, bone resorption.

Despite the histopathologic similarities between gingivitis and periodontitis, evidence is lacking that would indicate that periodontitis is an inevitable consequence of gingivitis. Furthermore, the pathogenic mechanisms explaining the progression of gingivitis lesions to periodontitis lesions are not clear, and the factors that lead to the initiation of periodontitis lesions are unknown. Clinical models of disease activity in periodontitis range from a continuous progression of disease during which loss of attachment occurs at a slow rate over long periods of time to

an episodic burst model in which loss of attachment occurs relatively rapidly during short periods of disease activity. Signature 35-37 Clinical data indicate that either mechanism could be operant in different patients or at different sites or at different times within the same patient, implying that the pathogenesis of periodontal attachment loss could differ between patients and sites and times. Understanding the pathologic mechanisms involved still awaits measurement methods that clearly differentiate between active and quiescent disease.

Bacterial Virulence

It is widely accepted that the initiation and progression of periodontitis are dependent upon the presence of microorganisms capable of causing disease. Although more than 300 species of microorganisms have been isolated from periodontal pockets, it is likely that only a small percentage of these are etiologic agents.³⁸ Among the characteristics that implicate an organism or group of organisms as etiologic agents are bacterial virulence factors. These are bacterial constituents or metabolites capable of either causing disruption of homeostatic or protective host mechanisms or causing the progression or initiation of the disease. If such bacterial virulence characteristics are truly contributing to disease pathogenesis, modification of such virulence factors should result in an improvement in clinical condition. Thus, the pathogenesis of periodontal disease lesions is in part dependent upon the virulence as well as the presence and concentrations of microorganisms capable of producing disease.

At least 3 characteristics of periodontal microorganisms have been identified that can contribute to their ability to act as pathogens: the capacity to colonize, the ability to evade antibacterial host defense mechanisms, and the ability to produce substances that can directly initiate tissue destruction. It is now apparent that within a given pathogenic species, such as Actinobacilius actinomycetemcomitans or Porphyromonas gingivalis, only a subset of bacterial types or clonal or genetic subtypes may be pathogenic.^{39,40} Thus the presence of a pathogenic bacterial species in the subgingival plaque may not by itself imply that a pathogen is present with virulence characteristics necessary to initiate or propagate periodontitis lesions. For example, recent data indicate that strains of A. actinomycetemcomitans in young patients with localized juvenile periodontitis differ from those in older patients with previously active disease in their ability to produce a leukotoxin that is thought to be an important virulence characteristic of this species.³⁹

Bacteria need to possess the ability to survive and propagate in periodontal pockets in the complex ecosystem of the biofilm. Some examples of factors that have been identified as promoting virulence of important periodontal pathogens follow. Virulent organisms can express appendages such as fimbriae or molecules such as adhesins which promote association with tissues or other bacteria. 41,42 Furthermore, virulence can be enhanced via the presence of a capsular polysaccharide (as in the case of P. ginqivalis) which provides resistance to host defenses such as antibody and complement. Some organisms are able to invade into or through host tissues, thereby creating a sequestered environment for their protection and gaining more direct access to susceptible host tissues. Two major periodontal disease pathogens, A. actinomycetemcomitans and P. gingivalis, are able to invade into the tissues. A. actinomucetemcomitans can pass through epithelial cells into the underlying connective tissues, 43 while P. gingivalis can invade and persist in epithelial cells. 44,45 It is likely that tissue invasiveness of these organisms may explain the difficulty in eradicating A. actinomycetemcomitans by mechanical root debridement, and could also explain the relatively high concentrations of serum antibody reactive with these two species in comparison with other bacteria in dental plaque.

An important feature of nearly all pathogenic microorganisms is the ability to evade the host defense mechanisms that would ordinarily control such infections and prevent disease. Foremost among these defense mechanisms in the periodontium is clearance of bacteria by neutrophils with the assistance of antibodies and complement proteins. In health, neutrophils appear to form a barrier at the plaque-tissue interface, controlling bacterial numbers and preventing ingress of bacteria or their products to the tissue surface. The immune system typically assists the neutrophil by producing antibody molecules that opsonize bacteria; such opsonic antibodies, alone or in concert with the comple-

ment system, allow the neutrophil to recognize, ingest, and degrade bacteria. The local repository of such antibody molecules is the gingival crevicular fluid (GCF), a modified inflammatory exudate which flows through the junctional and sulcular epithelium into the gingival crevice or pocket. Amongst a large variety of other molecules, the GCF contains serum components such as antibody molecules, 48 locally produced antibody molecules⁴⁹ and other substances, such as neutrophil granule constituents, 50,51 that can be reflective of local immunology and inflammatory processes. Antibacterial antibodies can provide many protective functions. Opsonic antibodies promote phagocytosis via interactions with phagocyte Fc receptors. 52-54 In some cases, antibodies can activate the complement system, an antibacterial cascade of naturally occurring proteins, which can deposit additional opsonins on the bacterial surface, release chemical mediators that recruit additional neutrophils, and deposit macromolecular complexes into the bacterial surface that will lyse and kill certain bacteria. Antibodies may also be produced that will specifically neutralize bacterial toxins and enzymes. 48,55 or that will disrupt bacterial colonization by preventing adherence to the tooth or epithelial surface or to other bacteria.56

Little is known about the sequence of events leading to the initial breakdown of this barrier and subsequent initiation of periodontitis. A great deal is known, however, about the mechanisms evolved by some periodontal bacteria to overcome this protective mechanism, and some examples of this are given below. Some organisms, such as strains of A. actinomycetemcomitans ⁵⁷ or Campylobacter rectus, ⁵⁸ produce leukotoxins that can kill neutrophils directly, thus disrupting the primary antibacterial defense mechanism in the gingival crevice. Secondly, some bacteria, such as P. gingivalis, produce proteolytic enzymes that either directly degrade antibody and complement proteins in the surrounding serum or GCF or prevent the accumulation of these molecules on the bacterial surface. 55,59 This activity would prevent accumulation of complement-derived chemotactic factors which would ordinarily recruit many additional neutrophils to the site of infection, as well as retard the phagocytosis of both the proteolytic bacteria themselves and other bacteria that are in close proximity. Third, some bacteria such as A. actinomycetemcomitans produce factors that suppress the immune response to itself and other bacteria, 60, 61 thereby diminishing the production of otherwise protective antibodies. Finally, as mentioned above, some bacteria can invade tissue cells and avoid contact with neutrophils and molecules of the immune system. Thus, pathogenic bacteria appear to have devised a number of means by which they can evade control by neutrophils, either by directly decreasing their numbers or by destroying host mechanisms meant to promote opsonization, phagocytosis, and bacterial killing.

The interaction between neutrophils, antibody, and complement provides primary protection against the deleterious effects of periodontal pathogens. In general, high levels of antibody do not appear in a patient's serum or GCF until some time after the disease process has initiated. High levels of antibodies reactive with bacterial virulence factors such as A. actinomucetemcomitans leukotoxin or P. gingivalis proteases, or with whole bacterial antigen preparations, do not occur until relatively late in the disease process and probably do not play an important role in prevention of disease initiation.^{48,62} However, it appears that in the case of the antibody response to A. actinomycetemcomitans and P. gingivalis in early-onset periodontitis patients the extent and severity of disease is the least in patients with the highest titers; thus, some antibody responses to periodontal disease pathogens may ultimately prevent or delay progression of existing disease.63,64

Destruction of Periodontal Tissues

The protective responses to periodontal pathogens may be overcome in a number of ways as outlined above, and the concentration of pathogens in subgingival plaque may reach a critical level required for initiation or progression of tissue destruction. Although at least two pathogenic bacteria have been shown to invade the superficial layers of the periodontal tissues, it is readily apparent from histologic observation that pathologic effects on connective tissue and alveolar bone occur at sites deep to the subgingival plaque and invading microorganisms. For this reason, in addition to the possible direct pathologic effects of bacteria on the periodontal tissues, it is clear that damage to the periodontium must also occur by indirect means. Bacterial

products must gain access to the cellular constituents of the gingival tissues and activate cellular processes that are destructive to collagenous connective tissue and bone.

Direct effects of bacteria. It is likely that direct pathological effects of bacteria and their products on the periodontium are significant during early stages of disease. Analysis of plague samples from patients with increasingly severe levels of gingival inflammation reveals a succession of bacterial species with increased capacity to directly induce an inflammatory response. For example, increased and persistent levels of Fusobacterium nucleatum in sites of mild gingivitis and the consequent production of its metabolic by-products may directly affect the gingival vasculature. The resulting edema and increase in production of GCF may provide the environment and nutrients that allow putative pathogens to flourish.³⁸ Although it is unknown whether or not gingivitis is a prerequisite to development of a periodontitis lesion, it is reasonable that the alteration of the gingival environment by toxic or proinflammatory by-products of the gingivitis flora can set the stage for increased concentrations of more virulent microorganisms within the plague mass.

It is also likely that bacteria can contribute to the pathogenesis of periodontal diseases directly by many other means. *P. gingivalis*, for example, is known to produce enzymes (proteases, collagenase, fibrinolysin, phospholipase A) that could directly degrade surrounding tissues in the superficial layers of the periodontium. In addition it produces metabolic by-products such as H₂S, NH₃, and fatty acids that are toxic to surrounding cells. 45,65-67 Furthermore, bacterial constituents such as lipopolysaccharide (LPS) are capable of inducing bone resorption in vitro. 68

Indirect effects of bacteria. Once the major protective elements in the periodontium have been overwhelmed by bacterial virulence mechanisms, a number of host-mediated destructive processes are initiated. Polymorphonuclear leukocytes (PMNs), which normally provide protection, can themselves contribute to tissue pathology. During the process of phagocytosis, these cells typically "spill" some of their enzyme content extracellularly during a process known as degranulation; some of these enzymes are capable of degrading the surrounding host tissues, namely collagen and basement membrane constituents, contributing to tissue damage.

There is increasing evidence that the bulk of tissue destruction in established periodontitis lesions is a result of the mobilization of the host tissues via activation of monocytes, lymphocytes, fibroblasts, and other host cells. Engagement of these cellular elements by bacterial factors, in particular bacterial lipopolysaccharide (LPS), is thought to stimulate production of both catabolic cytokines and inflammatory mediators including arachidonic acid metabolites such as prostaglandin $\rm E_2$ (PGE₂). Such cytokines and inflammatory mediators in turn promote the release of tissue-derived enzymes, the matrix metalloproteinases, which are destructive to the extracellular matrix and bone. 69,70

Once defensive mechanisms have been averted, the subgingival bacterial microflora has established itself as a predominantly anaerobic, Gram-negative infection. The pathologic appearance of the periodontitis lesion and the mediators, mediator precursors, and mRNA protein templates recognizable either in the GCF or within cellular elements of the gingival tissues are consistent with the expected outcome of a local infection with Gram-negative bacteria. Cytokines, molecules which are released by host cells into the local environment, provide molecular signals to other cells thereby affecting their function. Many cytokines are produced by cells in periodontitis lesions. Among the cytokines and inflammatory mediators most consistently found to be associated with periodontitis are the follow-

1. Interleukin 1 $(IL-1)^{71}$ is a pro-inflammatory, multifunctional cytokine, which among its many biological activities enables ingress of inflammatory cells into sites of infection, promotes bone resorption, stimulates eicosanoid (specifically, PGE₂) release by monocytes and fibroblasts, stimulates release of matrix metalloproteinases that degrade proteins of the extracellular matrix, and participates in many aspects of the immune response. IL-1 levels in general are elevated in both tissues^{72,73} and GCF⁷⁴⁻⁷⁷ from diseased, inflamed periodontal tissues compared to healthier sites, and elevated levels have been shown to be associated with active disease in animal models. 78 The predominant form in the periodontal tissues is IL-1 α , which is produced primarily by macrophages. 79,80

2. Interleukin 6 (IL-6)⁸¹ is a cytokine that stimulates plasma cell proliferation and therefore

antibody production and is produced by lymphocytes, monocytes, and fibroblasts. ⁸⁰ Levels of IL-6 have been shown to be elevated in inflamed tissues, higher in periodontitis than in gingivitis tissues, and higher in GCF from refractory periodontitis patients. ⁸²⁻⁸⁴ IL-6 has also been shown to stimulate osteoclast formation. Thus, this cytokine may in large part account for both the predominance of plasma cells in periodontitis lesions as well as bone resorption.

- 3. Interleukin 8 (IL-8)⁸⁵ is a chemoattractant that is mainly produced by monocytes in response to LPS, IL-1, or tumor necrosis factor alpha (TNF- α). It is present at high levels in periodontitis lesions, mainly associated with the junctional epithelium and macrophages,^{86,87} and its levels in GCF are higher in periodontitis patients than in healthy controls.⁸⁸ In addition to serving as a chemoattractant for neutrophils, it appears to selectively stimulate matrix metalloproteinase (MMP) activity from these cells, thus in part accounting for collagen destruction within periodontitis lesions.
- 4. Tumor necrosis factor alpha (TNF- α)^{89,90} shares many of its biological activities (pro-inflammatory properties, matrix metalloproteinase [MMP] stimulation, eiscosanoid production, and bone resorption) with IL-1. In addition, its secretion by monocytes and fibroblasts is stimulated by bacterial LPS.
- 5. Prostaglandin E_2 (PGE₂), 91,92 a vasoactive eicosanoid produced by monocytes and fibroblasts, induces bone resorption and MMP secretion. Many studies have shown the association of elevated levels of PGE₂ in tissues and GCF with periodontal inflammation, progressive periodontitis, and high-risk periodontitis patients (e.g., early-onset periodontitis, refractory periodontitis, diabetes mellitus). $^{93-100}$ The likely importance of eicosanoids in periodontal disease pathogenesis is underscored in several studies demonstrating the beneficial effects of both systemic and topical non-steroidal anti-inflammatory drugs on periodontitis in both animal models and in humans. $^{91,101-105}$

In summary, a simplified model for pathogenesis of periodontitis within the local lesion is the following: virulent microorganisms capable of initiating or propagating periodontal attachment loss must be present in the local lesion at a critical minimal infective dose. In susceptible individuals, or in susceptible periodontal sites within

susceptible individuals, protective mechanisms are breached exposing the underlying tissues and cells to bacterial components. Consequently, cellular components, including monocytes and fibroblasts, are stimulated by bacterial components such as LPS to produce many or all of the cytokines described above. These cytokines are capable of acting alone, or in concert, to stimulate inflammatory responses and catabolic processes such as bone resorption and collagen destruction via the MMPs.

Genetic Factors Promoting Periodontitis

As in any infectious disease, host susceptibility plays a major role in determining whether or not the presence of an infectious agent will ultimately lead to expression of disease or progression of preexisting disease. Genetic risk, one aspect of such host susceptibility, has been and is being examined. A summary of these data for specific periodontal diseases, appears below.

Adult periodontitis. Studies of adult periodontitis and periodontal health in twins have demonstrated that heredity accounts for a significant proportion of the population variance in various measures of periodontal diseases, such as gingival inflammation, probing depth, and radiographic bone levels. 106-108 Recent data indicate that a genetic variation or polymorphism in the gene encoding IL-1 (see above) is associated with severity of, and likely susceptibility to, periodontitis. 109 These polymorphisms are variations in the DNA sequence of genes coding for IL-1 α (the IL-1A gene) and IL-1B (the IL-1B gene). In a population of adult, non-smoking subjects of Caucasian Northern European heritage, a higher percentage of individuals with severe periodontal destruction tested positive for one of the genetic forms (alleles) of the IL-1A gene plus one of the Il-1B alleles more frequently than did subjects with less severe disease. Furthermore, one of the two alleles associated with risk for periodontitis is also known to be associated with elevated production of IL-1B, thus providing a possible biological explanation for the enhanced susceptibility of patient with this genotype for periodontitis.

Early-onset periodontitis: localized juvenile periodontitis (LJP), generalized juvenile periodontitis (GJP), rapidly progressive periodontitis (RPP). These diseases are characterized by their age of onset (usually post-pubertal), by the extent and severity of disease, by their oftentimes characteristic bacterial microflora, and to a

lesser extent by associated pathological and immunological characteristics. 110 These postpubertal forms of EOP have a familial distribution, 111-114 and a number of clinical and biological characteristics of EOP, including the epidemiology and immunologic responses, appear to be strongly influenced by race. 115-117 These data imply that it is possible that risk for EOP may be genetic. Although a number of genetic models have been tested using genetic segregation analysis, no consistent mode of inheritance for all forms of EOP has been observed. 118-124 One study has demonstrated genetic linkage of LJP with the Gc locus on chromosome 4 in one extended family, but this finding may not be generalizable to all families with EOP. 118,125

A number of hypotheses have been proposed implicating candidates for genetic risk factors. The observation that many patients with EOP, particularly LJP, have neutrophil chemotactic defects, point to factors related to neutrophil function such as receptors for chemotactic agents or molecules participating in signal transduction. 126-128 Associations of EOP with some antigens of the major histocompatibility complex (HLA) region have been demonstrated, indicating that heritable factors related to immunologic responsiveness may be associated with risk for EOP. 129 Additionally, poorly functional heritable forms of monocyte FcγRII, the receptor for human IgG2 antibodies, have been shown to be disproportionately present in patients with LJP. Such receptors cause monocytes to function poorly in phagocytosis of periodontal pathogens such as A. actinomycetemcomitans, because most of the antibody produced against this bacterium is of the IgG_2 subclass. ¹³⁰ Finally, studies have demonstrated hyperresponsiveness of monocytes from EOP patients with respect to their production of PGE₂ in response to LPS. This hyper-responsive phenotype could lead to increased connective tissue or bone loss due to inappropriately excessive production of these catabolic factors. 131,132

It is noteworthy that transmission of EOP in families, and many of the biologic characteristics of these diseases, may be explained by environmental factors as well as genetic factors, and some could be consequences of bacterial infection rather than the cause of such infections.

Pre-pubertal periodontitis. Prepubertal forms of periodontitis are usually subcategorized into a localized form (L-PP) and a generalized form (G-PP). L-PP is most commonly found in patients with no obvious health problems. Some, but not all, patients with L-PP display relative defects in neutrophil function and such patients can be frequently members of families in which other individuals have EOP. Additionally, it has been proposed that defects in cementum formation may predispose to L-PP.¹³³ In contrast, G-PP is frequently associated with systemic disorders that affect neutrophil function (chemotaxis, phagocytosis) or numbers. Among the disorders that can predispose to G-PP are leukocyte adhesion deficiencies (LAD), 134,135 a group of genetic disorders resulting in impaired adherence-dependent functions, as well as a number of other inherited phagocyte disorders (Chediak-Higashi syndrome, ¹³⁶ cyclic neutropenia, ¹³⁷ and Papillon-Lefevre syndrome ¹³⁸⁻¹⁴⁰), collagen defects (Ehler-Danlos syndrome type VIII¹⁴¹), and enzyme defects (acatalasia and hypophosphatasia 129,142-144). G-PP can, however, occur in patients with no such discernible defect: frequently, these patients are found in families of patients with other forms of early-onset periodontitis and thus may share common etiologic and pathogenic mechanisms with EOP.

Refractory periodontitis. This form of periodontitis is characterized by its relative resistance to repeated routine therapeutic attempts to control the progression of periodontal attachment loss. Studies have demonstrated that such patients, as seen in patients with EOP, can demonstrate hyperresponsive monocytic responses to bacterial LPS and produce high levels of PGE₂. ^{145,146} Some of these responses may be genetically determined in these patients.

SMOKING AND PATHOGENESIS OF PERIODONTAL DISEASE

It has been demonstrated that smoking is a risk factor for periodontitis in adults. The number of pack-years of exposure to tobacco smoke is associated with increased risk for adult periodontitis and increased disease severity in smokers compared to non-smokers. 147,148 Additionally, smoking has been shown to be associated with increased disease severity for the generalized forms of EOP (GJP, RPP). 149 The pathologic mechanisms proposed for the deleterious effects

of smoking on the periodontium include alterations of the periodontal tissue vasculature, direct alterative effects on the bacterial microflora, and inhibitory effects on immunoglobulin levels and antibody responses to plaque bacteria.

PERIODONTITIS ASSOCIATED WITH SYSTEMIC DISEASES

Many of the systemic conditions associated with or predisposing to periodontal attachment loss have as a common attribute defective neutrophil function. Severe periodontitis has been observed in primary neutrophil disorders including agranulocytosis, 150,151 cyclic neutropenia, 152,153 Chediak-Higashi syndrome, 136 and lazy leukocyte syndrome. 154 In addition, more frequent and severe periodontitis can be observed in many patients with diabetes mellitus, 148,155,156 Down's syndrome, 157,158 Papillon-Lefevre syndrome, 138-140 and inflammatory bowel disease, ^{128,159} which exhibit secondary neutrophil impairment. These disorders underscore the importance of the neutrophil in protection of the periodontium. It is assumed, though in nearly all cases not proven, that the pathogenic mechanisms leading to tissue destruction in patients with these diseases are similar to those in other forms of periodontitis as described above.

Unusual and severe forms of periodontitis can be more frequent in patients with certain severe combined and acquired immunodeficiency diseases. Furthermore, some patients with HIV infections develop necrotizing ulcerative periodontitis (NUP), in which acute destruction of the periodontium with bleeding, tissue necrosis, and pain can be observed. 18,20,160,161 It is important to note that this condition also occurs in the absence of HIV infection, and that its occurrence may be no more common than in the general population. 162 The pathogenesis of NUP associated with HIV infection is not clear; the subgingival bacterial flora in patients with HIV infections are not substantially different from that in other patients with periodontitis, with the exception that Candida and enteric pathogens can sometimes be found in some patients. Although it has been hypothesized that the dysregulation and suppression of the systemic and local immune response results in hyperresponsiveness of neutrophils in local lesions and exacerbation of the usual acute inflammatory response, 18,162 there are no definitive data to indicate that the pathogenesis of periodontal diseases in HIV-positive patients is different from that in HIV-negative patients.

A number of studies have demonstrated that there is a higher prevalence of periodontitis amongst patients with diabetes mellitus, and that diabetic patients have more severe periodontitis than do non-diabetic individuals. 148,156,163,164 Importantly, the degree of diabetic control and the duration of the disease are thought to be important factors contributing to the expression of periodontitis in diabetics. Additionally, the degree of control of periodontitis may influence metabolic control of diabetes mellitus. 165,166 Although the precise pathogenesis of periodontitis in such diabetic patients is not known, a number of pathologic features of this disease are consistent with increased risk for periodontitis. Factors such as impaired neutrophil function; microvascular alterations that could lead to impaired access of leukocytes and plasma proteins to the periodontium; and altered collagen metabolism reflective of increased collagenase activity, decreased collagen synthesis, and reduced bone matrix formation, all may contribute to the increased susceptibility of diabetics to periodontal breakdown.

SUMMARY

- 1. The initiation and propagation of most forms of gingivitis are dependent upon the presence and persistence of bacterial plaque. The histopathology of the gingivitis lesion and its stages are consistent with the following pathogenic mechanisms. Plaque bacteria contain or produce substances capable of causing inflammation. Such substances can have direct effects on the vasculature and on leukocytes, inducing vasodilatation, increased GCF flow, and emigration of neutrophils. Substances in bacterial plaque may also interact with host systems involved in inflammatory responses and thereby exacerbate clinical and histological parameters of inflammation. In more advanced stages of disease it is likely that bacterial antigens, via their ability to gain ingress to the periodontal tissues, activate host cells such as monocytes, lymphocytes, and fibroblasts, and thereby induce pathological changes that are consistent with a chronic inflammatory response.
- 2. Although a high proportion of sites that experience periodontal attachment loss display signs of gingival inflammation, there is little evidence demonstrating that gingivitis lesions will

always progress to become destructive periodontitis lesions. Furthermore, the pathologic processes that are operant during the initiation of attachment loss, whether alterations in the bacterial flora, fluctuations in host defense mechanisms, or other factors, are not well defined.

- 3. The pathology of periodontitis lesions are characteristic of, and consistent with, a subversion of host defenses against bacterial plaque pathogens and subsequent activation of bacterially-induced host-mediated processes that destroy periodontal tissues. Data indicate that pathogenic plaque bacteria have virulence characteristics that can prevent their efficient detection and elimination by the host, disable host cells and humoral factors, and directly adversely affect the tissues. The predominance of a Gramnegative bacterial flora, in combination with the cellular and cytokine profiles of the lesions, indicate the likelihood that bacterial LPS activation of monocytes and subsequent production of tissuedestructive cytokines is likely a major pathway for connective tissue attachment loss and bone loss in most forms of periodontitis. Such cytokines can cause tissue destruction via mobilization of tissue metalloproteinases, a major pathway for destruction of soft and hard connec-
- 4. Emerging data indicate that individual susceptibility to some forms of periodontal disease may be heritable. However, no definitive data in this regard are available. On the other hand, many inherited and acquired diseases characterized by diminished protective function of inflammatory and immunologic pathways are associated with more severe periodontal disease.

ACKNOWLEDGMENTS

The primary author of this revised paper is Dr. Harvey Schenkein. Members of the 1997-1998 Committee on Research, Science and Therapy included Drs. David L. Cochran, Chair; Thomas E. Van Dyke, Vice Chair; Timothy Blieden; Robert E. Cohen; William W. Hallmon; James E. Hinrichs; Angelo Mariotti; Leslie A. Raulin; Martha J. Somerman; Robert J. Genco, Consultant; Gary Greenstein, Board Liaison; Vincent J. Iacono, Board Liaison.

REFERENCES

1. The American Academy of Periodontology. Proceedings of the World Workshop in Clinical Periodontics. Chicago: The American Academy

- of Periodontology; 1989:I23-I24.
- 2. Armitage GC. Periodontal diseases: Diagnosis. *Ann Periodontol* 1996;1:37-215.
- 3. Consensus Report on Periodontal Diseases: Epidemiology and Diagnosis. *Ann Periodontol* 1996;1:216-222.
- 4. Cimasoni G. Crevicular fluid updated. *Monogr Oral Sci* 1983;12:III-VII 1-152.
- Greenstein G. The role of bleeding upon probing in the diagnosis of periodontal disease. A literature review. *J Periodontol* 1984;55:684-688.
- Löe H, Theilade E, Jensen SB. Experimental gingivitis in man. J Periodontol 1965;36:177-187.
- 7. Theilade E, Wright WH, Jensen SB, Löe H. Experimental gingivitis in man. II. A longitudinal clinical and bacteriological investigation. *J Periodont Res* 1966;1:1-13.
- 8. Page R. Gingivitis. J Clin Periodontol 1986;13:345-59.
- 9. Page RC, Schroeder H. *Periodontitis in Man and Other Animals. A Comparative Review.* Basel and New York: S. Karger; 1982.
- Payne WA, Page RC, Ogilvie AL, Hall WB. Histopathologic features of the initial and early stages of experimental gingivitis in man. *J* Periodont Res 1975;10:51-64.
- 11. Attström R, Egelberg J. Emigration of blood neutrophils and monocytes into the gingival crevices. *J Periodont Res* 1970;5:48-55.
- 12. Hellden L, Lindhe J. Enhanced emigration of crevicular leukocytes mediated by factors in human dental plaque. *Scand J Dent Res* 1973;81:123-129.
- 13. Johnson BD, Engel D. Acute necrotizing ulcerative gingivitis A review of diagnosis, etiology, and treatment. *J Periodontol* 1986;57:141-150.
- 14. Melnick SL, Roseman JM, Engel D, Cogen R. Epidemiology of acute necrotizing ulcerative gingivitis. *Epidemiol Rev* 1988;10:191-211.
- 15. Loesche WJ, Syed SA, Laughon BE, Stoll J. The bacteriology of acute necrotizing ulcerative gingivitis. *J Periodontol* 1982;53:223-230.
- Listgarten MA. Electron microscopic observations on the bacterial flora of acute necrotizing ulcerative gingivitis. *J Periodontol* 1965;36:328-339.
- 17. Silverman S Jr., Migliorati CA, Lozada NF, Greenspan D, Conant MA. Oral findings in people with or at high risk for AIDS: a study of 375 homosexual males. *J Am Dent Assoc* 1987;112:187-192.
- 18. Murray PA. HIV disease as a risk factor for periodontal disease. *Compendium Cont Educ Dent* 1994;15:1052, 1054-63.
- 19. Greenberg M. HIV-associated lesions. *Dermatol Clin* 1996;14:319-326.
- 20. Greenspan JS. Periodontal complications of HIV infection. *Compendium Cont Educ Dent* 1994;18:S694-S698.
- 21. Williams R. Periodontal disease [see comments]. *N Engl J Med* 1990;322:373-382.
- 22. Genco RJ, Zambon JJ, Christersson LA. The ori-

- gin of periodontal infections. Adv Dent Res 1988;2:245-59.
- 23. Genco RJ, Slots J. Host responses in periodontal diseases. *J Dent Res* 1984:63:441-451.
- 24. Löe H, Silness J. Periodontal disease in pregnancy. *Acta Odontol Scand* 1963; 21:533-551.
- Cohen DW, Shapiro J, Friedman L, Kyle GC, Franklin S. A longitudinal investigation of the periodontal changes during pregnancy and fifteen months postpartum: II. *J Periodontol* 1971;42:653-657.
- 26. Sooriyamoorthy M, Gower DB. Hormonal influences on gingival tissue: Relationship to periodontal disease. *J Clin Periodontol* 1989;16:201-208.
- Kornman KS. Age, supragingival plaque, and steroid hormones as ecological determinants of the subgingival flora, in host-parasite interactions in periodontal diseases. ASM, Washington 1982;132-138.
- Hassell TM, Page RC, Narayanon AS, Cooper CG. Diphenylhydantoin (chlantin) gingival hyperplasia. Drug induced abnormality of connective tissue. *Proc Natl Acad Sci* (USA) 1976;73:2909-2912.
- 29. Hassell TM, Hefti AF. Drug-induced gingival overgrowth: Old problem, new problem. *Crit Rev Oral Biol Med* 1991;2:103-137.
- 30. Seymour RA, Thomason JM, Ellis JS. The pathogenesis of drug-induced gingival overgrowth. *J Clin Periodontol* 1996;23:165-175.
- 31. Seymour RA, Jacobs DJ. Cyclosporin and the gingival tissues. *J Clin Periodontol* 1992;19:1-11.
- 32. Wynn RL. Update on calcium channel blockerinduced gingival hyperplasia. *Gen Dent* 1995;43:218,220,222.
- 33. Harel-Raviv M, Eckler M, Lalani K, Raviv E, Gornitsky M. Nifedipine-induced gingival hyperplasia. A comprehensive review and analysis. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1995;79:715-722.
- 34. Listgarten MA. Pathogenesis of periodontitis. *J Clin Periodontol* 1986;13:418-430.
- 35. Jeffcoat MK, Reddy MS. Progression of probing attachment loss in adult periodontitis. *J Periodontol* 1991;62:185-189.
- 36. Reddy MS, Jeffcoat MK. Periodontal disease progression. *Curr Opin Periodontol.* 1993:52-59.
- 37. Socransky SS, Haffajee AD, Goodson JM, Lindhe J. New concepts of destructive periodontal disease. *J Clin Periodontol* 1984;11:21-32.
- 38. Moore WEC, Moore LVH. The bacteria of periodontal diseases. *Periodontol 2000* 1994;5:66-77.
- 39. Zambon JJ, Haraszthy VI, Hariharan G, Lally ET, Demuth DR. The microbiology of early-onset periodontitis: Association of highly toxic *Actinobacillus actinomycetemcomitans* strains with localized juvenile periodontitis. *J Periodontol* 1996; 67:282-290.
- Haffajee AD, Socransky SS. Microbial etiological agents of destructive periodontal diseases.

- Periodontol 2000 1994;5:78-111.
- 41. Hamada S, Fujiwara T, Morishima S, et al. Molecular and immunological characterization of the fimbriae of *Porphyromonas gingivalis*. *Microbiol Immunol* 1994;38:921-930.
- 42. Wilson M, Henderson B. Virulence factors of *Actinobacillus actinomycetemcomitans* relevant to the pathogenesis of inflammatory periodontal diseases. *FEMS Microbiol Rev* 1995;17:365-379.
- Fives-Taylor P, Meyer D, Mintz K. Characteristics of Actinobacillus actinomycetemcomitans invasion of and adhesion to cultured epithelial cells. Adv Dent Res 1995:9:55-62.
- 44. Lamont RJ, Chan A, Belton CM, Izutsu K, Vasel D, Weinberg A. *Porphyromonas gingivalis* invasion of gingival epithehal cells. *Infect Immun* 1995;63:3878-3885.
- 45. Holt SC, Bramanti TE. Factors in virulence expression and their role in periodontal disease pathogenesis. *Crit Rev Oral Biol Med* 1991;2:177-281.
- 46. Hart TC, Shapira L, Van Dyke TE. Neutrophil defects as risk factors for periodontal diseases. *J Periodontol* 1994;65:521-529.
- 47. Miyasaki KT. The neutrophil: Mechanisms of controlling periodontal bacteria. *J Periodontol* 1991;62:761-774.
- 48. Ebersole JL. Systemic humoral immune responses in periodontal disease. *Crit Rev Oral Biol Med* 1990;1:283-331.
- 49. Tew JC, Marshall DC, Burmeister JA, Ranney R. Relationship between gingival crevicular fluid and serum antibody titers in young adults with generalized and localized periodontitis. *Infect Immun* 1985;49:487-493.
- Lamster IB, Novak MJ. Host mediators in gingival crevicular fluid: Implications for the pathogenesis of periodontal disease. Crit Rev Oral Biol Med 1992;3:31-60.
- 51. Lamster IB. The host response in gingival crevicular fluid: Potential applications in periodontitis clinical trials. *J Periodontol* 1992;63:1117-1123.
- 52. Wilson ME, Bronson PM, Hamilton RG. Immunoglobulin G₂ antibodies promote neutrophil killing of *Actinobacillus actinomycetem-comitans*. *Infect Immun* 1995;63:1070-1075.
- Baker PJ, Wilson ME. Opsonic IgG antibody against Actinobacillus actinomycetemcomitans in localized juvenile periodontitis. Oral Microbiol Immunol 1989;4:98-105.
- 54. Wilson ME, Genco RJ. The role of antibody, complement and neutrophils in host defense against *Actinobacillus actinomycetemcomitans*. *Immunol Invest* 1989;18:187-209.
- 55. Cutler CN, Arnold RR, Schenkein HA. Inhibition of C3 and IgG proteolysis enhances phagocytosis of *Porphyromonas gingivalis*. *J Immunol* 1993;151:7016-7029.
- 56. Saito A, Hosaka Y, Nakagawa T, et al. Significance of serum antibody against surface antigens of *Actinobacillus actinomycetemcomitans* in patients with adult periodontitis. *Oral Microbiol Immunol* 1993;8:146-153.

- 57. Tsai CC, McArthur WP, Baehni PC, Hammond B, Taichman N. Extraction and partial characterization of a leukotoxin from a plaque-derived Gram-negative microorganism. *Infect Immun* 1979; 25:427-439.
- 58. Gillespie MJ, Smutko J, Haraszthy GG, Zambon JJ. Isolation and partial characterization of the *Campylobacter rectus* cytotoxin. *Microb Pathog* 1993; 14: 203-215.
- 59. Schenkein HA. Failure of *Bacteroides gingivalis* W83 to accumulate bound C3 following opsonization with serum. *J Periodont Res* 1989;24:20-27.
- 60. Shenker BJ, Vitale LA, Welham DA. Immune suppression induced by *Actinobacillus actinomycetemcomitans*: Effects on immunoglobulin production by human B cells. *Infect Immun* 1990;58:3856-3862.
- 61. Shenker BJ, Tsai CC, Taichman NS. Suppression of lymphocyte responses by Actinobacillus actinomycetemcomitans. J Periodont Res 1982;17:462-465.
- Haffajee AD, Socransky SS, Taubman MA, Sisson J, Smith DJ. Patterns of antibody response in subjects with periodontitis. *Oral Microbiol Immunol* 1995;10:129-137.
- 63. Califano JV, Gunsolley JC, Nakashima K, Schenkein HA, Wilson ME, Tew JG. Influence of anti-Actinobacillus actinomycetemcomitans Y4 (serotype b) LPS on severity of generalized early-onset periodontitis. Infect Immun 1996:64:3908-3910.
- 64. Gunsolley JC, Burmeister JA, Tew JG, Best AM, Ranney RR. Relationship of serum antibody to attachment level patterns in young adults with juvenile periodontitis or generalized severe periodontitis. J Periodontol 1987;58:314-320.
- 65. Birkedal-Hansen H, Taylor RE, Zambon JJ, Barwa PK, Neiders ME. Characterization of collagenolytic activity from strains of *Bacteroides gingivalis*. *J Periodont Res* 1988;23:258-264.
- 66. Singer RE, Buckner BA. Butyrate and propionate: Important components of toxic dental plaque extracts. *Infect Immun* 1981;32:458-463.
- 67. Robertson PB, Lantz M, Marucha PT, Kornman KS, Trummel CL, Holt SC. Collagenolytic activity associated with *Bacteroides* species and *Actinobacillus actinomycetemcomitans*. *J Periodont Res* 1982;17:275-283.
- 68. Hausmann E, Raisz LG, Miller WA. Endotoxin: Stimulation of bone resorption in tissue culture. *Science* 1970;168:862-864.
- 69. Offenbacher S. Periodontal diseases: Pathogenesis. *Ann Periodontol* 1996;1:821-878.
- 70. Birkedal-Hansen H. Role of cytokines and inflammatory mediators in tissue destruction. *J Periodont Res* 1993;28:500-510.
- 71. Tatakis DN. Interleukin-1 and bone metabolism: A review. *J Periodontol* 1993;64:416-31.
- 72. Jandinski JJ, Stashenko P, Feder LS, et al. Localization of interleukin-1 beta in human periodontal tissues. *J Periodontol* 1991;62:36-43.
- 73. Stashenko P, Fujiyoshi P, Obernesser MS,

- Prostak L, Haffajee AD, Socransky SS. Levels of interleukin-1 beta in tissue from sites of active periodontal disease. *J Clin Periodontol* 1991:18:548-554.
- 74. Preiss DS, Meyle J. Interleukin-1 beta concentration of gingival crevicular fluid. *J Periodontol* 1994;65:423-428.
- 75. Wilton JM, Bampton JL, Griffiths GS, et al. Interleukin-1 beta (Il-1ß) levels in gingival crevicular fluid from adults with previous evidence of destructive periodontitis. A cross-sectional study. *J Clin Periodontol* 1992;19:53-57.
- Yavuzyilmaz E, Yamalik N, Bulut S, Ozen S, Ersoy F, Saatci U. The gingival crevicular fluid interleukin-1 beta and tumour necrosis factoralpha levels in patients with rapidly progressive periodontitis. *Aust Dent J* 1995;40:46-49.
- 77. Hou LT, Liu CM, Rossomando EF. Crevicular interleukin-1 beta in moderate and severe periodontitis patients and the effect of phase I periodontal treatment. *J Clin Periodontol* 1995; 22:162-167.
- 78. Smith MA, Braswell LD, Collins JG, et al. Changes in inflammatory mediators in experimental periodontitis in the rhesus monkey. *Infect Immun* 1993;61:1453-1459.
- Matsuki Y, Yamamoto T, Hara K. Interleukin-1 mRNA-expressing macrophages in human chronically inflamed gingival tissues. Am J Pathol 1991;138:1299-1305.
- Matsuki Y, Yamamoto T, Hara K. Detection of inflammatory cytokine messenger RNA (mRNA)-expressing cells in human inflamed gingiva by combined in situ hybridization and immunohistochemistry. *Immunol* 1992;76:42-47.
- 81. Lotz M. Interleukin-6: A comprehensive review. Cancer Treat Res 1995:80:209-233.
- Yamazaki K, Nakajima T, Gemmell E, Polak B, Seymour G, Hara K. IL-4- and IL-6-producing cells in human periodontal disease tissue. *J Oral Pathol Med* 1994;23:347-353.
- 83. Reinhardt RA, Masada MP, Kaldahl WB, et al. Gingival fluid IL-1 and IL-6 levels in refractory periodontitis. *J Clin Periodontol* 1993;20:225-231.
- 84. Geivelis M, Turner DW, Pederson ED, Lamberts B. Measurements of interleukin-6 in gingival crevicular fluid from adults with destructive periodontal disease. *J Periodontol* 1993;64:980-983.
- 85. Bickel M. The role of interleukin-8 in inflammation and mechanisms of regulation. *J Periodontol* 1993;64:456-460.
- Tonetti MS, Imboden MA, Gerber L, Lang N, Laissue J, Mueller C. Localized expression of mRNA for phagocyte-specific chemotactic cytokines in human periodontal infections. *Infect Immun* 1994;62:4005-4014.
- 87. Fitzgerald JE, Kreutzer DL. Localization of interleukin-8 in human gingival tissues. *Oral Microbiol Immunol* 1995;10:297-303.
- 88. Tsai CC, Ho YP, Chen CC. Levels of interleukin-1 beta and interleukin-8 in gingival crevicular fluids in adult periodontitis. *J Periodontol*

- 1995;66:852-859.
- 89. Rink L, Kirchner H. Recent progress in the tumor necrosis factor-alpha field. *Int Arch Allergy Immunol* 1996;111:199-209.
- 90. Moldawer LL. Biology of proinflammatory cytokines and their antagonists. *Crit Care Med* 1994;22:S3-S7.
- Offenbacher S, Heasman PA, Collins JG. Modulation of host PGE₂ secretion as a determinant of periodontal disease expression. J Periodontol 1993;64:432-444.
- 92. Offenbacher S, Collins JG, Heasman PA. Diagnostic potential of host response mediators. *Adv Dent Res* 1993;7:175-181.
- Zhou J, Zou S, Zhao W, Zhao Y. Prostaglandin E₂ level in gingival crevicular fluid and its relation to the periodontal pocket depth in patients with periodontitis. *Chin Med Sci J* 1994;9:52-55.
- 94. Heasman PA, Collins JG, Offenbacher S. Changes in crevicular fluid levels of interleukin-1 beta, leukotriene B4, prostaglandin E₂, thromboxane B2 and tumour necrosis factor alpha in experimental gingivitis in humans. *J Periodont Res* 1993;28:241-247.
- 95. Sengupta S, Fine J, Wu-Wang C et al. The relationship of prostaglandins to CAMP, IgG, IgM and alpha-2-macroglobulin in gingival crevicular fluid in chronic adult periodontitis. *Arch Oral Biol* 1990;35:593-596.
- 96. Offenbacher S, Odle BM, Van Dyke TE. The use of crevicular fluid prostaglandin E₂ levels as a predictor of periodontal attachment loss. *J Periodont Res* 1986;21:101-112.
- 97. Offenbacher S, Odle BM, Gray RC, Van Dyke TE. Crevicular fluid prostaglandin E levels as a measure of the periodontal disease status of adult and juvenile periodontitis patients. *J Periodont Res* 1984;19:1-13.
- 98. Offenbacher S, Farr DH, Goodson JM. Measurement of prostaglandin E in crevicular fluid. *J Clin Periodontol* 1981;8:359-367.
- 99. Goodson JM, Dewhirst FE, Brunetti A. Prostaglandin E₂ levels and human periodontal disease. *Prostaglandins* 1974;6:81-85.
- 100. Ohm K, Albers HK, Lisboa BP. Measurement of eight prostaglandins in human gingival and periodontal disease using high pressure liquid chromatography and radioimmunoassay. *J Periodont Res* 1984;19:501-511.
- Paquette D. Potential role of nonsteroidal antiinflammatory drugs in the treatment of periodontitis. Compendium Cont Educ Dent 1992;13:1174-1179.
- Howell TH, Williams RC. Nonsteroidal antiinflammatory drugs as inhibitors of periodontal disease progression. Crit Rev Oral Biol Med 1993;4:177-196.
- 103. Offenbacher S, Williams RC, Jeffcoat MK, et al. Effects of NSAIDs on beagle crevicular cyclooxygenase metabolites and periodontal bone loss. *J Periodont Res* 1992;27:207-213.
- Heasman PA, Seymour RA. The effect of a systemically-administered nonsteroidal anti-inflam-

- matory drug (flurbiprofen) on experimental gingivitis in humans. *J Clin Periodontol* 1989:16:551-556.
- 105. Williams RC, Jeffcoat MK, Howell TH, et al. Altering the progression of human alveolar bone loss with the non-steroidal anti-inflammatory drug flurbiprofen. J Periodontol 1989;60:485-490.
- 106. Michalowicz BS, Aeppli DP, Virag JG, et al. Periodontal findings in adult twins. *J Periodontol* 1991;62:293-299.
- 107. Michalowicz BS, Aeppli DP, Kuba RK, et al. A twin study of genetic variation in proportional radiographic alveolar bone height. *J Dent Res* 1991;70:1431-1435.
- 108. Michalowicz BS. Genetic and heritable risk factors in periodontal disease. J Periodontol 1994;65:479-488.
- 109. Kornman KS, Crane A, Wang H-Y, et al. The interleukin-1 genotype as a severity factor in adult periodontal disease. *J Clin Periodontol* 1997;24:72-77.
- Schenkein HA, Van Dyke TE. Early-onset periodontitis: Systemic aspects of etiology and pathogenesis. *Periodontol* 2000 1994;6:7-25.
- 111. Benjamin SD, Baer PN. Familial patterns of advanced alveolar bone loss in adolescence (periodontosis). *Periodontics* 1967;5:82-88.
- 112. Butler JH. A familial pattern of juvenile periodontitis (periodontosis). *J Periodont* 1969;40:115-118.
- 113. Page RC, Vandesteen GE, Ebersole JL, Williams BL, Dixon IL, Altman LC. Clinical and laboratory studies of a family with a high prevalence of juvenile periodontitis. *J Periodontol* 1985; 56:602-610.
- 114. Sussman HI, Baer PN. Three generations of periodontosis: Case reports. *Ann Dent* 1978;7:8-11.
- 115. Gunsolley JC, Tew JG, Gooss CM, Burmeister JA, Schenkein HA. Effects of race and periodontal status on antibody reactive with Actinobacillus actinomycetemcomitans strain Y4. J Periodont Res 1988;23:303-307.
- 116. Gunsolley JC, Tew JG, Conner T, Burmeister JA, Schenkein HA. Relationship between race and antibody reactive with periodontitis-associated bacteria. *J Periodont Res* 1991;26:59-63.
- 117. Löe H, Brown LJ. Early-onset periodontitis in the United States of America. *J Periodontol* 1991;62:608-616.
- 118. Boughman JA, Halloran SL, Roulston D, et al. An autosomal-dominant form of juvenile periodontitis: Its localization to chromosome 4 and linkage to dentinogenesis imperfecta and Gc. *J Craniofac Genet Dev Biol* 1986;6:341-350.
- 119. Beaty TH, Boughman JA, Yang P, Astemborski J, Suzuki J. Genetic analysis of juvenile periodontitis in families ascertained through an affected proband. Am J Hum Genet 1987;40:443-452.
- 120. Hart TC, Marazita ML, Schenkein HA, Diehl S. Re-interpretation of the evidence for X-linked dominant inheritance of juvenile peri-

- odontitis. J Periodontol 1992;63:169-173.
- 121. Long JC, Nance WE, Waring P, Burmeister JA, Ranney RR. Early onset periodontitis: A comparison and evaluation of two proposed modes of inheritance. *Genet Epidemiol* 1987;4:13-24.
- 122. Marazita ML, Burmeister JA, Gunsolley JC, Koertge TE, Lake K, Schenkein HA. Evidence for autosomal dominant inheritance and racespecific heterogeneity in early onset periodontitis. J Periodontol 1994;65:623-630.
- 123. Saxén L, Nevanlinna HR. Autosomal recessive inheritance of juvenile periodontitis: Test of a hypothesis. *Clin Genet* 1984;25:332-335.
- 124. Saxén L. Heredity of juvenile periodontitis. *J Clin Periodontol* 1980;7:276-288.
- 125. Hart TC, Marazita ML McCanna KM, Schenkein HA, Diehl S. Reevaluation of the chromosome 4q candidate region for early onset periodontitis. *Hum Genet* 1993;91:416-422.
- 126. Van Dyke TE, Schweinebraten M, Cianciola LJ, Offenbacher S, Genco RJ. Neutrophil chemotaxis in families with localized juvenile periodontitis. *J Periodont Res* 1985;20:503-514.
- 127. Van Dyke TE, Warbington M, Gardner M. Offenbacher S. Neutrophil surface protein markers as indicators of defective chemotaids in LJP. *J Periodontol* 1990;61:180-184.
- 128. Van Dyke TE, Vaikuntam J. Neutrophil function and dysfunction in periodontal disease. Curr Opin Periodontol 1994;1:19-27.
- 129. Sofaer JA. Genetic approaches in the study of periodontal diseases. *J Clin Periodontol* 1990;17:401-408.
- Wilson ME, Kalmar JR. FcγRIIa (CD32): A potential marker defining susceptibility to localized juvenile periodontitis. *J Periodontol* 1996;67:323-331.
- 131. Shapira L, Soskolne WA, Sela MN, Offenbacher S, Barak V. The secretion of PGE₂, IL-1 beta, IL-6, and TNF alpha by adherent mononuclear cells from early onset periodontitis patients. *J Periodontol* 1994;65:139-146.
- 132. Shapira L, Soskolne W, Van Dyke TE. Prostaglandin E₂ secretion, cell maturation, and CD14 expression by monocyte-derived macrophages from localized juvenile periodontitis patients. *J Periodontol* 1996;67:224-228.
- 133. Watanabe K. Prepubertal periodontitis: A review of diagnostic criteria, pathogenesis, and differential diagnosis. *J Periodont Res* 1990;25:31-48.
- 134. Anderson DC; Schmalsteig FC; Finegold MJ, et al. The severe and moderate phenotypes of heritable Mac-1, LFA-1 deficiency: Their quantitative definitions and relation to leukocyte dysfunction and clinical features. *J Infect Dis* 1985;152:668-689.
- 135. Waldrop TC, Anderson DC, Hallmon WW, Schmalstieg FC, Jacobs RL. Periodontal manifestations of the heritable Mac-1, LFA-1, deficiency syndrome. Clinical histopathologic and molecular characteristics. J Periodontol 1987;58:400-416.
- 136. Hamilton RE, Giansanti JS. The Chediak-

- Higashi syndrome: Report of a case and review of the literature. *Oral Surg Oral Med Oral Pathol* 1974;37:754-761.
- 137. Cohen DW, Morris AL. Periodontal manifestations of cyclic neutropenia. *J Periodontol* 1961;32:159-168.
- 138. Dekker G, Jansen LH. Periodontosis in a child with hyperkeratosis palmoplantaris. J Periodontol 1958;29:266-271.
- 139. Galanter DR, Bradford S. Case report. Hyperkeratosis palmoplantaris and periodontosis: The Papillon-Lefevre syndrome. *J Periodontol* 1969:40:40-46.
- 140. Ingle JL. Papillon-Lefevre syndrome: Precocious periodontosis with associated epidermal lesions. *J Periodontol* 1959;30:230-237.
- 141. Hart TC. Genetic risk factors for early-onset periodontitis. *J Periodontol* 1996;67:355-366.
- 142. Casson MH. Oral manifestations of primary hypophosphatasia. A case report. *Br Dent J* 1969;127:561-566.
- 143. Bruckner RJ, Rickles NH, Porter DR. Hypophosphatasia with premature shedding of teeth and aplasia of cementum. *Oral Surg Oral Med Oral Pathol* 1962;15:1351-1369.
- 144. Baer PN, Brown NC, Hammer JE, III. Hypophosphatasia: Report of two cases with dental findings. *Periodontics* 1964;2:209-215.
- 145. Garrison SW, Nichols FG. LPS-elicited secretary responses in monocytes: Altered release of PGE₂ but not IL-1 beta in patients with adult periodontitis. *J Periodont Res* 1989;24:88-95.
- 146. Payne JB, Peluso JF, Jr., Nichols FC. Longitudinal evaluation of peripheral blood monocyte secretory function in periodontitisresistant and periodontitis-susceptible patients. *Arch Oral Biol* 1993;38:309-317.
- 147. Genco RJ. Assessment of risk of periodontal disease. Compendium Cont Educ Dent 1994; (Suppl 18):S678-683.
- 148. Grossi SG, Zambon JJ, Ho AW, et al. Assessment of risk for periodontal disease. I. Risk indicators for attachment loss. *J Periodontol* 1994;65:260-267.
- 149. Schenkein HA, Gunsolley JC, Koertge TE, Schenkein JG, Tew JG. Assessment of the effects of smoking on the extent and severity of early-onset periodontitis. *J Am Dent Assoc* 1995;126:1107-1113.
- 150. Saglam F, Atamer T, Onan U, Soydinc M, Kirac K. Infantile genetic agranulocytosis (Kostmann type). A case report. *J Periodontol* 1995;66:808-810.
- 151. Lamster IB, Oshrain RL, Harper DS. Infantile agranulocytosis with survival into adolescence: Periodontal manifestations and laboratory findings. A case report. *J Periodontol* 1987;58:34-39
- Rylander H, Ericsson I. Manifestations and treatment of periodontal disease in a patient suffering from cyclic neutropenia. *J Clin Periodontol* 1981;8:77-87.
- 153. Prichard JF, Ferguson DM, Windmiller J, Hurt W.

- Prepubertal periodontitis affecting the deciduous and permanent dentition in a patient with cyclic neutropenia. A case report and discussion. *J Periodontol* 1984;55:114-122.
- 154. Miller ME, Oski FA, Harris MB. Lazy leukocyte syndrome: A new disorder of neutrophil function. *Lancet* 1971;1:665-669.
- 155. The American Academy of Periodontology. Diabetes and periodontal diseases (Position Paper). *J Periodontol* 1996;67:166-176.
- 156. Shlossman M. Diabetes mellitus and periodontal disease—a current perspective. Compendium Cont Educ Dent 1994;15:1018,1020-1024 passim.
- 157. Izumi Y, Sugiyama S, Shinozuka O, Yamazaki T, Ohyama T, Ishikawa I. Defective neutrophil chemotaxis in Down's syndrome patients and its relationship to periodontal destruction. *J Periodontol* 1989;60:238-242.
- Shaw L, Saxby MS. Periodontal destruction in Down's syndrome and in juvenile periodontitis. J Periodontol 1986;57:709-715.
- 159. Vrotsos JA, Vrahopoulos TP. Effects of systemic diseases on the periodontium. *Curr Opin Periodontol* 1996;3:19-26.
- 160. Barr CE. Periodontal problems related to HIV-1 infection. *Adv Dent Res* 1995;9:147-151.
- Horning GM, Cohen ME. Necrotizing ulcerative gingivitis, periodontitis, and stomatitis: Clinical staging and predisposing factors. *J Periodontol* 1995;66:990-998.
- Ryder MI. Periodontal considerations in the patient with HIV. Curr Opin Periodontol 1993:43-51.
- 163. Emrich L, Shlossman M, Genco RJ. Periodontal disease in non-insulin-dependent diabetes mellitus. *J Periodontol* 1991;62:123-131.
- 164. Pinson M, Hoffman WH, Garnick JJ, Litaker M. Periodontal disease and type I diabetes mellitus in children and adolescents. J Clin Periodontol 1995;22:118-23.
- 165. Miller LS, Manwell MA, Newbold D et al. The relationship between reduction in periodontal inflammation and diabetes control: A report of 9 cases. *J Periodontol* 1992;63:843-848.
- 166. Aldridge JP, Lester V, Watts TL, Collins A, Viberti G, Wilson R. Single-blind studies of the effects of improved periodontal health on metabolic control in type 1 diabetes mellitus. J Clin Periodontol 1995;22:271-275.

Individual copies of this informational paper may be obtained by contacting the Science and Education Department at The American Academy of Periodontology, Suite 800, 737 N. Michigan Avenue, Chicago, IL 60611-2690; voice: 312/573-3230; fax: 312/573-3234; e-mail: adriana@perio.org. Members of The American Academy of Periodontology have permission of the Academy, as copyright holder, to reproduce up to 150 copies of this document for not-for-profit, educational purposes only. For information on reproduction of the document for any other use or distribution, please contact Rita Shafer at the Academy Central Office; voice: 312/573-3221; fax: 312/573-3225; e-mail: rita@perio.org.